

Research Journal of Pharmaceutical, Biological and Chemical Sciences

Inhibition Of Uric Acid Formation By *Mimosa pudica* L. Herb Extract Tablets.

Sri Adi Sumiwi^{1*}, Marline Abdassah², Raisa Muthiarani³, Kirthika Gopal Krishnan⁴,
Restri Akhsanitami⁵, Ade Zuhrotun⁶, Jutti Levita⁷

^{1,3,4,5,7}Department of Pharmacology and Clinical Pharmacy, Faculty of Pharmacy, Universitas Padjadjaran, Indonesia

²Department of Pharmaceutical, Faculty of Pharmacy, Universitas Padjadjaran, Indonesia

⁶Department of Pharmaceutical Biology, Faculty of Pharmacy, Universitas Padjadjaran, Indonesia

ABSTRACT

Mimosa pudica Linn has been known to have various pharmacological effects. Among these are, as antidiabetic activity, antitoxin, antihepatotoksin antiinflammatory, antioxidant, wound healing, analgesic, and blood diseases (S.Varnika et. Al, 2012). Extracts and fractions with concentrations of 250, 125 and 62.5 μ L effective inhibit of xanthine oxidase. The ethanol extract had the highest activity. Tablet formulation of the ethanol extract of *Mimosa pudica* Linn. herb. In this work we developed *Mimosa pudica* L. tablets using Vivapur[®]PH102 and studied the inhibitory activity of the tablets on uric acid formation. Extraction herbaceous *Mimosa pudica* Linn and evaluation of the active ingredient, drying viscous *Mimosa pudica* Linn extract. The tablet formulation. additives used are AEROSIL200 and VIVAPUR[®]PH102 than as a binder and filler are also crusher, primojel is functioning as a disintegrator, combination with talc, magnesium stearate and AEROSIL200 also is functioning as absorbent. inhibitory activity assay of the tablets on uric acid formation was performed in vitro by measuring uric acid absorbance at 292 nm. Ex vivo study was performed by measuring the concentration of uric acid in oxonic calcium and chicken liver juice induced-mice blood after the mice were treated with *Mimosa pudica* L. extract tablets. Allopurinol was used as drug control. *Mimosa pudica* L. extract tablets containing the highest Vivapur[®]PH102 (86%) showed the shortest disintegration time (1 minute 40 seconds). The tablets inhibited uric acid formation (IC₅₀ of *Mimosa pudica* tablet = 5.381 ppm, IC₅₀ of the extract = 3.5202 ppm). Its inhibitory activity is weaker than allopurinol (IC₅₀ of allopurinol = 2.181 ppm). Ex vivo study showed that *Mimosa pudica* L. extract tablets 125 mg/kg of body weight reduced 36% of uric acid level in hyper-urisemic mice ($\alpha=0.05$). *Mimosa pudica* tablet 125 mg/kg of body weight inhibited uric acid formation in hyper-uricemic mice, therefore this pharmaceutical dosage form could be proposed as anti-hyperurisemic drug.

Keywords: allopurinol, gout, hyperurisemic, xanthine, xanthine oxidase, *Mimosa pudica* Linn

*Corresponding author

INTRODUCTION

Mimosa pudica L. (Family Mimosaceae or Fabaceae), known as *putri malu* in Indonesia or lajvanthi or chuimui in India [1], has been popular lately and invited many researchers to explore its pharmacological activities. This plant contain mimosine, adrenalin-like substance, crocetin dimethyl ester, tubuline and a new class phytohormone turgorines, quercetin-7-rhamnoside, luteolin-glycoside, acacetin-7-rutinoside, tannin up to 10 %, d-xylose, d-glucuronic acid, mimosainic acid, mimosinamine, green yellow fatty oil up to 17 % and many other chemical constituents [2, 3]. *M. pudica* has been proven to show anti-ulcer [4], anti-inflammatory [5], anti-microbial (in combination with *Tridax procumbens*) [6], anti-toxin against *Naja naja* and *Bangarus caeruleus* venoms [7], wound healing [8], anti-convulsant [9], anti-oxidant [10], anthelmintic [11], aphrodisiac [12], hepatoprotector [13, 14], and xanthine oxidase inhibitory activities [15].

Xanthine oxidase (XO) is an enzyme that catalyses the oxidation of xanthine and hypoxanthine into uric acid. According to the work of Oettl and Reibnegger who studied the inhibitory action of pteridine and its analogues on XO, the most characteristic features of an inhibitor are aromaticity and no substitution at position 7 of the pteridine ring [16]. XO inhibitors which block the terminal step in uric acid biosynthesis, can lower the plasma level of uric acid hence are generally employed for the treatment of gout, as proven by Ishibuchi and colleagues, who studied the structure and activity relationships of 1-phenylpyrazoles as XO inhibitors [17].

Studies of the pharmacological activities of *M. pudica* were mainly performed on the extracts, whilst the pharmacological activity of this plant's pharmaceutical product has not been provided. Therefore, in this work we manufactured *M. pudica* L. tablets using Vivapur®PH102 and studied the inhibitory activity of the tablets on uric acid formation.

MATERIALS AND METHODS

Plant materials:

Dried leaves of *M. pudica* L., were extracted 3 x 24h using ethanol 70%. The extracts were collected and the solvent was evaporated in a water-bath apparatus until a viscous extract was obtained.

Experimental animals:

The experimental animals used were male rats (*Rattus norvegicus*) weighing 150-250 g obtained from rats were quarantined for one week and fasted for 18-20 hours before blood sampling, with water provided *ad libitum*. The experiment was approved by the Health Research Ethics Committee of the Faculty of Medicine, Universitas Padjadjaran No: 265/NN6C 132/KEPK/PN/2015

Chemical materials:

ethanol 70% (BrataChem-Indonesia), Vivapur®PH102, sodium starch glycolate (Primojel®), magnesium stearate, aerosil®, dimethylsulfoxide, xanthine oxidase assay kit (Worthington Biochemical Corporation), Allopurinol tablets.

Instruments:

Spectroscopy UV-Vis

Methods:

Procedure 1:

Formulation:

Six formulas for direct tableting (omitting granulation) using various percentages of Vivapur®PH102 were showed in Table 1.

Table 1. Formulas of *M. pudica* L. Tablets

Formula	A	B	C	D	E	F
<i>M. pudica</i> extract	11.5%	11.5%	11.5%	11.5%	11.5%	11.5%
Vivapur®PH102	81.0%	82.0%	83.0%	84.0%	85.0%	86.0%
Primojel	5.0%	4.0%	3.0%	2.0%	1.0%	--
Magnesium stearate	1.0%	1.0%	1.0%	1.0%	1.0%	1.0%
Talc	1.0%	1.0%	1.0%	1.0%	1.0%	1.0%
Aerosil	0.5%	0.5%	0.5%	0.5%	0.5%	0.5%

In vitro study: This study was performed on the best-evaluated tablets. 20 tablets were weighed and powdered. 50 mg of the powder (equals to 11.5 mg of the extract) was added with 3 drops of DMSO and phosphate buffer to the final volume of 100 mL. This solution was diluted into 5 concentrations: 11.50; 23.00; 34.50; 46.00; 57.50 ppm. To 1 mL of each solution was added 1.9 mL of phosphate buffer 50 mM pH 7.5 and 1 mL of xanthine solution 0.15 mM. The mixture was added with 0.1 mL of xanthine oxidase and was measured its absorbance at 292 nm. The same procedure was carried out on the extract and the standard drug (allopurinol).

Ex vivo study:

25 Swiss-Webster mice (*Mus musculus*) were treated according to the Ethics Approval Committee. Their blood was taken to measure the uric acid level for baseline. The mice were kept fasted (only drink-water was allowed to be given) for 18 h then were separated into 5 groups:

- I. PGA 2% only
- II. This group was induced to hyper-uricemic with calcium oxonic 300 mg/kg of body weight and chicken liver juice 0.5 mL/25 g of body weight. This group was used as negative-control.
- III. This group was induced to hyper-uricemic with calcium oxonic 300 mg/kg of body weight and chicken liver juice 0.5 mL/25 g of body weight, and treated with allopurinol 13 mg/kg of body weight. This group was used as positive-control.
- IV. This group was induced to hyper-uricemic with calcium oxonic 300 mg/kg of body weight and chicken liver juice 0.5 mL/25 g of body weight, and treated with *M. pudica* extract 125 mg/kg of body weight
- V. This group was induced to hyper-uricemic with calcium oxonic 300 mg/kg of body weight and chicken liver juice 0.5 mL/25 g of body weight, and treated with *M. pudica* tablet 125 mg/kg of body weight

Their blood was taken during 1, 2, 3, 4, and 5 hours after treatment, and were measured using UA Sure®. The inhibition percentage of xanthine oxidase activity was calculated according to the formula:

$$(A_{\text{control}} - A_{\text{sample}}) / A_{\text{control}} \times 100\% \text{ [18].}$$

Data Analysis

Statistical analysis was performed with Anova method with SPSS software tools.

hypothesis:

H0: $\tau_1 = \tau_2 = \dots = \tau_4 = 0$ (There are 4 treatment on the formation of uric acid)

H0: At least one sign $\neq 0$ (There is a difference of 4 treatment on the formation of uric acid).

Tabel 4 Varians of Analysis

Diversity	Sum of Squares	Db	Central Square	F	Sig,
Treatment	11727,987	3	3909,329	630,837	0,000
Block (Concentration)	131,116	4	32,779	5,289	0,011
Error	74,365	12	6,197		
Total	11933,467	19			

According to the table F the F value [0.05: 3: 12] = 3.49. In a test analysis of variance, reject the hypothesis if $F_{count} > F_{table} (\alpha: k, (n-k))$ or if the value $sig < \alpha$, in other cases accept the null hypothesis. Therefore the value of F (630 837) > F table (3.49) and sig (0000) < α (0.05), the null hypothesis is rejected. So it can be concluded that there 4 treatment is difference on the inhibition of the formation of uric acid.

Duncan test

Based on testing Anava Table 4, resulted in the value of H0 is rejected, we can do further testing to determine which treatment gives different effect on the inhibition of the formation urat. Results test is as follows:

Tabel 5 Duncan Test Advanced Treatment

Treatment	Subset		
	1	2	3
Alopurinol	53,1264		
<i>Mimosa pudica</i> extract Tablet	57,2866		
<i>Mimosa pudica</i> extract	59,2594		
Negative Control	112,247		

The results shows that the difference in treatment effect on the inhibition of the formation of uric acid can be divided into three groups: the first group is a positive control, the second group is extracts tablet and extract and third group is a negative control

RESULTS AND DISCUSSION

Table.2 Granule and tablet evaluation

Evaluation parameters	A	B	C	D	E	F
Water content (%)	2.10	2.60	3.10	3.90	4.10	4.90
Compressibility (%)	16.95	17.73	20.30	21.22	24.32	26.66
Density (g/mL)	1.79	1.62	1.54	1.50	1.44	1.42
Flow rate (g/sec) (with vibration)	6.86	6.76	6.48	6.49	6.48	6.50
Repose angle (°) (without vibration)	-	-	-	-	21.51°	20.65°

Repose angle (°) (with vibration)	19.61 ^a	16.18 ^a	26.04 ^a	25.54 ^a	25.07 ^a	28.14 ^a
Weight uniformity (mg)	499.33	497.14	498.49	499.88	498.65	497.03
Thickness (mm)	4.35	4.40	4.52	4.60	4.91	5.34
Diameter size (mm)	12.10	12.10	12.10	12.10	12.10	12.00
Hardness (N)	82.45	83.00	84.40	85.40	86.50	84.20
Friability (%)	1.02	0.71	0.42	0.33	0.14	0.13
Disintegration (minute)	4.40	3.48	3.30	3.40	3.50	1.40

Table 2 showed that all formulas fulfill USP requirements. Furthermore we calculated the correlation between Vivapur®PH102 concentration and disintegration time (Fig.1). An increasing of Vivapur®PH102 and decreasing of Primojel in a mixture of tablet, resulted a lower crushing strength, shorter disintegration time.

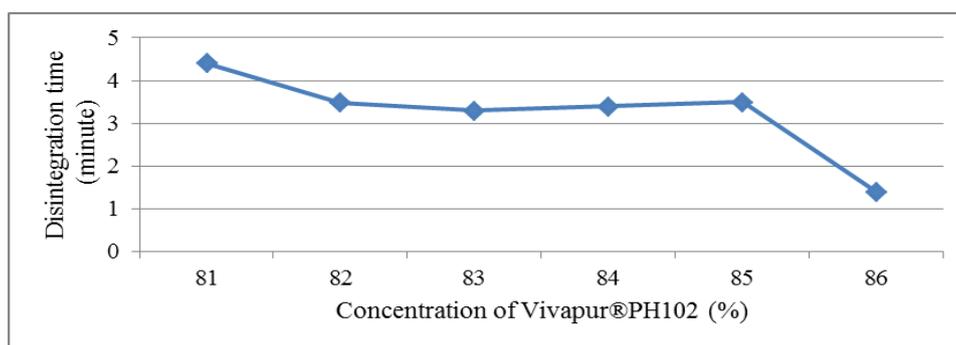


Fig.1 Correlation between Vivapur®PH102 concentration and disintegration time

This result was compared to the work of Lahdenpää et al (1997) who concluded that the increasing of Avicel® PH-102 granular and especially Avicel® PH-200 in the mixture of tablet, resulted a lower crushing strength, shorter disintegration time, and smaller weight variation [19].

The result of *in vitro* study could be seen in Fig.2 showed the inhibition percentage of uric acid formation by *M. pudica* tablet (blue), *M. pudica* extract (red), and allopurinol (green). Trendline and data labels were provided for the tablet, extract, and drug control in different colours. The data showed that IC₅₀ of *M. pudica* tablet is 68.04 ppm, IC₅₀ of the extract is 32.75 ppm, whilst IC₅₀ of allopurinol is 18.73 ppm.

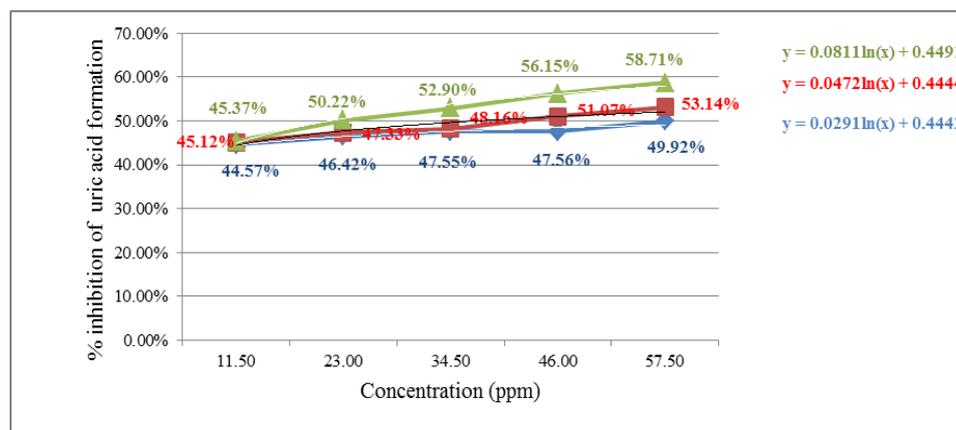


Fig.2 Inhibition percentage of uric acid formation by *M. pudica* tablet (blue), *M. pudica* extract (red), and allopurinol (green)

This data proved that direct tableting of *M. pudica* extract decreases the activity to 48.13% . Although previous study of Marczyński concluded that direct tableting, omitting granulation, prevented biological activity loss of active substances [20].

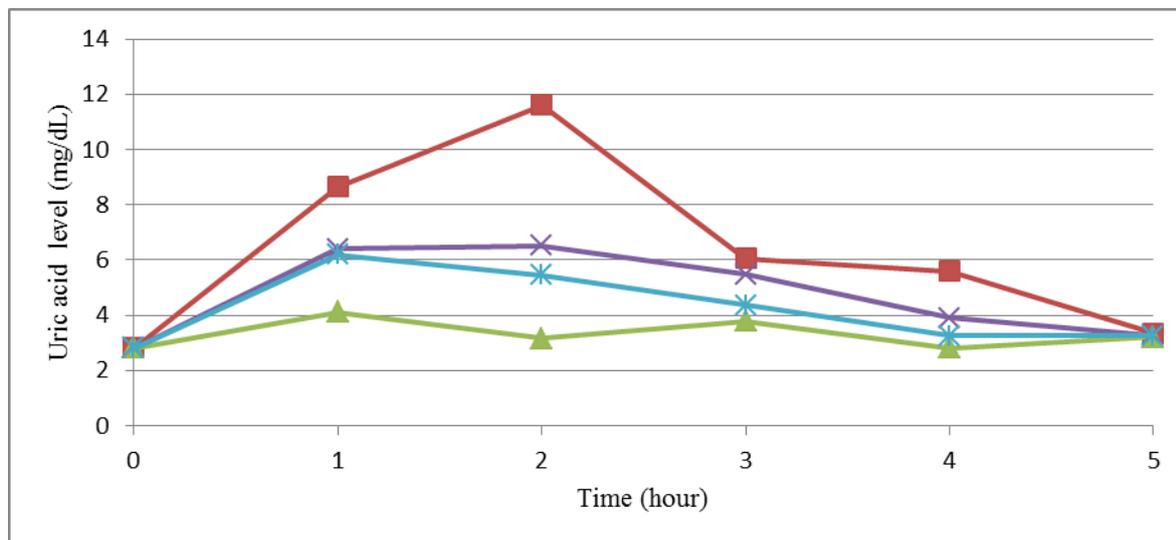


Fig.3 Inhibitory activity on uric acid formation by *Mimosa pudica* tablet (purple), extract (blue) compared to negative control (red) and allopurinol (green)

The result of *ex vivo* study could be seen in Fig.3. Fig.3 showed the inhibitory activity on uric acid formation by *Mimosa pudica* tablet and extract. The result showed that *Mimosa pudica* L. tablet 125 mg/kg of body weight and extract reduced 36% and 43%, respectively, of uric acid level in hyper-uricemic mice ($\alpha=0.05$).

Results

Table 1 Average concentrations of uric acid for Five Minutes

Treatment	Uric acid concentration (units / mL)
Alopurinol	57,287
<i>Mimosa pudica</i> extract	59,259
Tablet	53,126
Mimosa pudica extract	112,247
Negative Control	57,287
Alopurinol	57,287

CONCLUSIONS

Mimosa pudica L. herb extract tablet 125 mg/kg of body weight inhibited uric acid formation in hyper-uricemic mice, therefore this pharmaceutical dosage form could be proposed as anti-hyperuricemic drug.

REFERENCES

- [1] Chauhan BS, Johnson DE. 2009. Germination, emergence and dormancy of *Mimosa pudica*. *Weed Biology and Management*. 9(1): 38-45
- [2] Joseph B, George J, Mohan J. 2013. Pharmacology and traditional uses of *Mimosa pudica*. *International Journal of Pharmaceutical Sciences and Drug Research*. 5(2): 41-44

- [3] Sanaye MM, Joglekar CS, Pagare NP. 2015. Mimososa-A brief overview. *Journal of Pharmacognosy and Phytochemistry*. 4(2): 182-187
- [4] Vinothapooshan Gv, Sundar K. 2010. Anti-ulcer activity of *Mimosa pudica* leaves against gastric ulcer in rats. *Research Journal of Pharmaceutical, Biological and Chemical Sciences* 1(4): 606-614
- [5] Mistry S, Patidar R, Vyas V, Jena J, Dutt KR. 2012. Anti-inflammatory activity of *Mimosa pudica* Linn. (Mimosaceae) leaves: An ethnopharmacological study. *J. Pharm. Sci. and Res.* 4(3): 1789-1791
- [6] Sharma MC, Sharma S. 2010. Phytochemical and pharmacological screening of combined *Mimosa pudica* Linn and *Tridax procumbens* for in vitro antimicrobial activity. *International Journal of Microbiological Research*. 1 (3): 171-174
- [7] Meenatchisundaram S, Priyagrace S, Vijayaraghavan R, Velmurugan A, Parameswari G, Michael A. 2009. Antitoxin activity of *Mimosa pudica* root extracts against *Naja naja* and *Bangarus caeruleus* venoms. *Bangladesh Journal of Pharmacology*. 4(2): 105-109
- [8] Kannan S, Jesuraj SAV, Kumar ESJ, Saminathan K, Suthakaran R, Kumar MR, Devi BP. 2009. Wound healing activity of *Mimosa pudica* Linn formulation. *Int. J. PharmTech. Research*. 1(4): 1554-1558
- [9] Ngo Bum E, Soudi S, Ayissi ER, Dong C, Lakuolo NH, Maidawa F, Seke PFE, Nanga LD, Taiwe GS, Dimo T, Njikam N, Rakotonirina A, Kamanyi A. 2011. Anxiolytic activity evaluation of four medicinal plants from Cameroon. *Afr. J. Tradit. Complement Altern. Med.* 8(5): 130-139
- [10] Patro G, Bhattamisra SK, Mohanty BK, Sahoo HB. 2016. *In vitro* and *in vivo* antioxidant evaluation and estimation of total phenolic, flavonoidal content of *Mimosa pudica* L. *Pharmacognosy Res.* 8(1): 22-28
- [11] Bendgude RD, Maniyar MG, Kondawar MS, Patil SB, Hirave RV. 2012. Anthelmintic activity of leaves of *Mimosa pudica*. *International Journal of Institutional Pharmacy and Life Sciences*. 2(1): 120-125
- [12] Pande M, Pathak A. 2009. Aphrodisiac activity of roots of *Mimosa pudica* Linn. ethanolic extract in mice. *International Journal of Pharmaceutical Sciences and Nanotechnology*. 2(1): 477-486
- [13] Rajendran R, Hemalatha S, Akasakalai K, MadhuKrishna CH, Sohil B, Sundaram V, Sundaram RM. 2009. Hepatoprotective activity of *Mimosa pudica* leaves against carbontetrachloride induced toxicity. *Journal of Natural Products*. 2: 116-122
- [14] Kumaresan R, Veerakumar S, Elango V. 2015. A study on hepatoprotective activity of *Mimosa pudica* in Albino rats. *International Journal of Pharmacognosy and Phytochemical Research*. 7(2): 337-339
- [15] Nguyen MTT, Awale S, Tezuka Y, Tran QL, Watanabe H, and Kadota S. 2004. Xanthine oxidase inhibitory activity of Vietnamese medicinal plants. *Biol. Pharm. Bull.* 27(9): 1414-1421
- [16] Oettl K and Reibnegger G. 1999. Pteridines as inhibitors of xanthine oxidase: structural requirements. *Biochimica et Biophysica Acta*. 1430(2): 387-395
- [17] Ishibuchi S, Morimoto H, Oe T, Ikebe T, Inoue H, Fukunari A, Kamezawa M, Yamada I, Naka Y. 2001. Synthesis and structure-activity relationships of 1-phenylpyrazoles as xanthine oxidase inhibitors. *Bioorg. Med. Chem. Lett.* 11(7): 879-882
- [18] Sahgal G, Ramanathan S, Sasidharan S, Mordi MN, Ismail S, Mansor SM. 2009. *In vitro* antioxidant and xanthine oxidase inhibitory activities of methanolic *Swietenia mahagoni* seed extracts. *Molecules*. 14: 4476-4485
- [19] Lahdenpää E, Niskanen M, Yliruusi J. 1997. Crushing strength, disintegration time and weight variation of tablets compressed from three Avicel® PH grades and their mixtures. *European Journal of Pharmaceutics and Biopharmaceutics*. 43(3): 315-322
- [20] Marczyński Z. 2009. Tableting technology of a dry extract from *Solidago virgaurea* L. with the use of silicified microcrystalline cellulose (Prosolv) and other selected auxiliary substances. [Polim Med.](#) 39(4): 51-60